Inflammation is a reflexive response to infection, the binding of antibodies to antigens within the body, mechanical irritation, or injury. Microbes that breach epithelial barriers, for instance, directly activate complement and toll-like receptors, two principal components of the innate immune system. The activation of these sentinels triggers the synthesis and release of inflammatory mediators with acute effects on the vasculature. Localized vasodilation, increased vascular permeability, extravasation of plasma (and humoral) proteins, and migration of leukocytes into the affected tissue produce the classic signs of inflammation: calor, dolor, rubor, tumor, and functio laesa. A positive feedback loop initiates the production of additional inflammatory cytokines once infiltrating leukocytes become activated. Anti-inflammatory homeostatic mechanisms reverse these processes as the infectious agent is cleared by the innate and adaptive immune systems. The hypothalamic–pituitary–adrenal axis and glucocorticoids in particular are essential in limiting and resolving the inflammatory process.

Whereas restricted inflammation is beneficial, excessive or persistent inflammation incites tissue destruction and disease. Together, inflammatory disorders such as allergies, asthma, autoimmune diseases, and sepsis are a major cause of illness and death. Asthma affects approximately 21.9 million adults and 8.9 million children in the United States alone. The prevalence of autoimmune diseases, which affect 8.5 million Americans, is also noteworthy. Rheumatoid arthritis, Graves’ disease, glomerulonephritis, type 1 diabetes mellitus, multiple sclerosis, thyroiditis, pernicious anemia, systemic lupus erythematosus, psoriasis, and vitiligo account for most of these autoimmune diseases. Sepsis is fatal for roughly 30 percent of the 700,000 patients affected annually in the United States. Glucocorticoids are indicated for the treatment of many of these diverse conditions. The efficacy of glucocorticoids in alleviating inflammatory disorders results from the pleiotropic effects of the glucocorticoid receptor on multiple signaling pathways. Pleiotropy can, however, also have adverse effects: growth retardation in children, immunosuppression, hypertension, inhibition of wound repair, osteoporosis, and metabolic disturbances. All these harmful properties contraindicate prolonged glucocorticoid therapy. Here, we review mechanisms whereby glucocorticoids inhibit inflammation and the therapeutic limitations of these hormones. We then provide a prospectus for research on drugs that dissociate the beneficial and detrimental effects of glucocorticoids.
hypothalamus to control the secretion of corticotropin-releasing hormone into the hypophyseal portal system (Fig. 1). In turn, corticotropin-releasing hormone stimulates the release of corticotropin from the anterior pituitary. Corticotropin induces the synthesis and secretion of cortisol by the adrenal cortex. Most of the secreted cortisol (approximately 90 percent) is bound to corticosteroid-binding globulins in the blood. Free cortisol is the biologically active form of the hormone and is converted to cortisone by type 2 11β-hydroxysteroid dehydrogenase. Conversely, type 1 11β-hydroxysteroid dehydrogenase converts cortisone into cortisol.

The glucocorticoid receptor is a member of the steroid-hormone–receptor family of proteins. It binds with high affinity to cortisol; the bound cortisol promotes the dissociation of molecular chaperones, including heat–shock proteins, from the receptor (Fig. 2). Within the cell, cortisol acts in three ways. First, the cortisol–glucocorticoid receptor complex moves to the nucleus, where it binds as a homodimer (see the Glossary) to DNA sequences called glucocorticoid-responsive elements. The resulting complex recruits either coactivator or corepressor proteins that modify the structure of chromatin, thereby facilitating or inhibiting assembly of the basal transcription machinery and the initiation of transcription by RNA polymerase II. This process is highly dynamic in cell culture and is presumably so in vivo. Second, regulation of other glucocorticoid-responsive genes involves interactions between the cortisol–glucocorticoid receptor complex and other transcription factors, such as nuclear factor-κB (NF-κB) (Fig. 2). These latter actions seem to occur at lower cortisol levels than the cortisol–glucocorticoid receptor–glucocorticoid-responsive element complex needs to change transcription. The third mechanism is glucocorticoid signaling through membrane-associated receptors and second messengers (so-called nongenomic pathways) (Fig. 2). Evidence indicates that the glucocorticoid receptor inhibits inflammation through all three mechanisms: direct and indirect genomic effects and nongenomic mechanisms.

**STRUCTURE OF THE GLUCOCORTICOID RECEPTOR**

The human glucocorticoid receptor (GR) gene is one locus on chromosome 5q31–32 (Fig. 3). Even so, variation in the structure and the expression of the gene generates diversity in glucocorticoid signaling. The genomic structure includes three transcription-initiation sites; each produces an alternative first exon that is spliced to a common exon 2 (Fig. 3). Though the first exon is not translated, there is a potential for functional differences among exons 1A, 1B, and 1C because dexamethasone up-regulates all three GR transcripts to a similar degree in acute lymphoblastic leukemia T cells but depresses these transcripts to different degrees in a B-cell line. These observations highlight the importance of understanding the regulation of the expression of the glucocorticoid receptor in health and disease.

Human GR messenger RNA (mRNA) has alternative splice variants. Where exons 2 through 8 are constant components of GR mRNA, there are two exon 9 isoforms that can be spliced to produce mature mRNA. Splicing of exon 9α produces GRα mRNA, which is translated into a protein with a unique sequence of 50 amino acids at its carboxy end (Fig. 3). The glucocorticoid receptor α isoform binds cortisol, DNA, and other transcription factors, thereby modifying transcriptional activity of target genes. Limited evidence suggests glucocorticoid receptor α may act through nongenomic pathways. Splicing of exon 9β produces GRβ mRNA, which is translated into a protein with 15 distinct amino acids at its carboxy end (Fig. 3). Although glucocorticoid receptor β protein forms homodimers that bind DNA, it does not bind any ligands examined so far and fails to activate transcription. Glucocorticoid receptor β can also form heterodimers with glucocorticoid receptor α and interfere with the function of this protein. The relative levels of glucocorticoid receptor α and β in a cell influence the cell’s sensitivity to glucocorticoid, with higher levels of glucocorticoid receptor β leading to glucocorticoid resistance. The inflammatory cytokines tumor necrosis factor α (TNF-α) and interleukin-1 can selectively up-regulate the levels of glucocorticoid receptor β, suggesting its role in inflammation.

Alternative translation-initiation sites within exon 2 produce additional isoforms of the glucocorticoid receptor (Fig. 3). Translation from the first methionine codon in GRα and GRβ mRNA produces proteins that consist of 777 amino acids (glucocorticoid receptor α-A) and 742 amino acids (glucocorticoid receptor β-A). Translation from a second methionine produces proteins with 751 ami-
Figure 1. Pathways of Communication among the Immune System, the Hypothalamic–Pituitary–Adrenal Axis, and Other Tissues Influenced by Immune Signals and Glucocorticoids.

The diagram also shows other important influences on the hypothalamic–pituitary–adrenal axis. Red lines denote inhibition, and blue and black arrows activation.
no acids (glucocorticoid receptor α-B) and 716 amino acids (glucocorticoid receptor β-B), respectively. There are important functional differences between the two isoforms: glucocorticoid receptor α-B has roughly twice the biologic activity of glucocorticoid receptor α-A in gene-expression studies in vitro.26 The finding that the two isoforms are expressed at different ratios in various types of cells and tissues also suggests that they may have distinct functions in vivo.27

POST-TRANSLATIONAL MODIFICATIONS OF THE GLUCOCORTICOID RECEPTOR

The human glucocorticoid receptor has five serine residues that are phosphorylated under different conditions by cyclin-dependent kinases and mitogen-activated protein kinases (MAPKs) (Fig. 3).28 The phosphorylation of several of the serines is dependent on the binding of ligands such as cortisol to the glucocorticoid receptor, whereas other serines are phosphorylated in a ligand-independent manner. The specific combination of serines that are phosphorylated has distinct effects on its transcriptional activity. For example, the glucocorticoid receptor is found primarily in the cytoplasm and is inactive when phosphorylated at serine 211.28 Another important modification of the glucocorticoid receptor that cortisol binding induces is the covalent attachment of ubiquitin to the receptor (Fig. 3), thus marking it for degradation by the proteasome;29 however, this process can be cell-type specific.30 Recent studies show that sumoylation (the attachment of small, ubiquitin-related modifiers) of the glucocorticoid receptor potentiates its transcriptional activity.31,32 Little is known about the effect of post-translational modifications on the repression of gene transcription, interactions with other transcription factors, or nongenomic signaling pathways.

NEUROENDOCRINE REGULATION OF INFLAMMATION

Interactions among the nervous system, the hypothalamic–pituitary–adrenal axis, and components of the innate and adaptive immune system play a key role in the regulation of inflammation and immunity (Fig. 1).2,8 For instance, cytokines and inflammatory mediators activate peripheral pain receptors whose axons project to the dorsal horn and synapse with the lemniscal tract, which in turn carries pain signals to the thalamus and the somatosensory cortex. Activation of this nociceptive pathway ultimately stimulates hypothalamic–pituitary–adrenal activity. Glucocorticoids inhibit the synthesis of cytokines and inflammatory mediators, thus forming a negative feedback loop. Cytokines can also act directly on the brain to activate the hypothalamic–pituitary–adrenal axis. Dysregulation of this neuroendocrine loop by hyperactivity or hypactivity of the hypothalamic–pituitary–adrenal axis causes systemic changes in inflammation and immunity.2,8

Hyperactivity of the hypothalamic–pituitary–adrenal axis in the absence of inflammation, as in Cushing’s syndrome, causes immunosuppression and increased susceptibility to infection.33 Physical pain, emotional trauma, and caloric restriction also activate the hypothalamic–pituitary–adrenal axis and cause immunosuppression.34,35 In contrast, decreased activity of the axis and low levels of glucocorticoids increase susceptibility to and the severity of inflammation. Patients with Addison’s disease, for example, require supplemental glucocorticoids during infection and inflammation to prevent the toxic effects of cytokines.36 Dysregulation of the hypothalamic–pituitary–adrenal axis by inflammation is associated with adverse outcomes among patients with the acute respiratory distress syndrome.37 Likewise, acquired glucocorticoid resistance is a common occurrence in patients with severe rheumatoid arthritis.38 Glucocorticoid resistance is a common finding and can be due to decreased expression of glucocorticoid receptor α, increased expression of glucocorticoid receptor β, or activation of MAPK, which phosphorylates the glucocorticoid receptor and thereby inhibits glucocorticoid signaling.39,40

Figure 2 (facing page). Three General Mechanisms of Action of Glucocorticoids and the Glucocorticoid Receptor in the Inhibition of Inflammation.

TNF-α denotes tumor-necrosis factor α, HSP heat-shock protein, mRNA messenger RNA, and P phosphate. The three mechanisms are nongenomic activation, DNA-dependent regulation, and protein interference mechanisms (e.g., NF-κB elements). Black arrows denote activation, the red line inhibition, the red dashed arrow repression, and the red X lack of product (i.e., no mRNA).
### Glossary

<table>
<thead>
<tr>
<th>Term</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Activator protein 1</td>
<td>An inflammatory transcription factor composed of two polypeptide subunits (c-Jun homodimers or c-Jun–Fos heterodimers)</td>
</tr>
<tr>
<td>Coactivator</td>
<td>A class of eukaryotic proteins that modulate the structure of chromatin or serve as a bridge between transcription factors and RNA polymerase II to facilitate the transcription of target genes</td>
</tr>
<tr>
<td>Corepressor</td>
<td>A class of eukaryotic proteins that modulate the structure of chromatin or inhibit RNA polymerase II to suppress transcription of target genes</td>
</tr>
<tr>
<td>Glucocorticoid-responsive element</td>
<td>A DNA motif usually found in the promoter of glucocorticoid-responsive genes that binds glucocorticoid receptor and enhances transcription</td>
</tr>
<tr>
<td>Heterodimer</td>
<td>A complex of two different protein chains held together by non-covalent bonds</td>
</tr>
<tr>
<td>Homodimer</td>
<td>A complex of two identical protein chains held together by non-covalent bonds</td>
</tr>
<tr>
<td>Negative glucocorticoid-responsive element</td>
<td>A DNA motif usually found in the promoter of glucocorticoid-responsive genes that binds glucocorticoid receptor and represses transcription</td>
</tr>
<tr>
<td>Nuclear factor-κB (NF-κB)</td>
<td>An inflammatory transcription factor composed of two polypeptides from the Rel family of proteins</td>
</tr>
<tr>
<td>Pleiotropy</td>
<td>The ability of one gene to have multiple phenotypic effects</td>
</tr>
<tr>
<td>Transcription</td>
<td>The synthesis of RNA from a DNA template</td>
</tr>
<tr>
<td>Transactivation</td>
<td>The stimulation of transcription by a specific protein (i.e., transcription factor)</td>
</tr>
<tr>
<td>Transrepression</td>
<td>The inhibition of transcription by a specific protein (i.e., transcription factor)</td>
</tr>
</tbody>
</table>

### Antiinflammatory Signaling Mechanisms

Glucocorticoids and the glucocorticoid receptor reside at the apex of a regulatory network that blocks several inflammatory pathways (Fig. 4). For example, glucocorticoids can inhibit prostaglandin production through three independent mechanisms: the induction and activation of annexin I, the induction of MAPK phosphatase 1, and the repression of transcription of cyclooxygenase 2. Annexin I (also called lipocortin-1) is an antiinflammatory protein that physically interacts with and inhibits cytosolic phospholipase A₂ (cPLA₂).\(^{41-44}\) This calcium-binding protein requires elevated calcium levels and phosphorylation by the protein kinases MAPK, calcium/calmodulin–dependent kinase II, and MAPK interacting kinase to exert its enzymatic activity.\(^{45}\) The activation of cPLA₂ by inflammatory stimuli begins with the movement of the phospholipase from the cytosol to the perinuclear membrane, where it hydrolyzes phospholipids containing arachidonic acid. Glucocorticoids induce annexin I, which by inhibiting cPLA₂, blocks the release of arachidonic acid and its subsequent conversion to eicosanoids (i.e., prostaglandins, thromboxanes, prostacyclins, and leukotrienes). Mice lacking annexin I have elevated levels of cPLA₂, an exaggerated inflammatory response, and partial resistance to the antiinflammatory action of glucocorticoids.\(^{46-48}\) A strong correlation exists between basal and corticotropic-stimulated cortisol levels and the expression of annexin I in neutrophils in humans, but the clinical importance of annexin I as an antiinflammatory protein is unknown.\(^{49}\)

A second antiinflammatory protein induced by glucocorticoids is MAPK phosphatase 1 (Fig. 4).\(^{50-52}\) Cytokines, bacterial and viral infections, and ultraviolet radiation are but a few of the inflammatory signals that activate MAPK cascades.\(^{16}\) Ultraviolet light triggers a kinase cascade that phosphorylates and activates Jun N-terminal kinase, which in turn phosphorylates the transcription factor c-Jun. Phosphorylated c-Jun homodimers and c-Jun–Fos heterodimers bind DNA sequences called activator protein 1 response elements and induce the transcription of inflammatory and immune genes.\(^{16}\) Glucocorticoid-induced MAPK phosphatase 1 dephosphorylates and inactivates Jun N-terminal kinase, thereby inhibiting c-Jun–mediated transcription. MAPK phosphatase 1 also dephosphorylates and inactivates all members of the MAPK family of proteins, including Jun N-terminal kinase, extracellular-signal–related kinase 1 and 2, and p38 kinase. Consequently, MAPK phosphatase 1 may also inhibit cPLA₂ activity by blocking its phosphorylation by MAPKs and MAPK-interacting kinase. In addition to blocking an essential upstream component of the c-Jun pathway, glucocorticoids and the glucocorticoid receptor directly interfere with c-Jun–mediated transcription (Fig. 4). Transcriptional interference between the glucocorticoid receptor and c-Jun homodimers (and c-Jun–Fos heterodimers) results from protein–protein interactions and has proved to be a major antiinflammatory mechanism.\(^{16}\)

The cortisol–glucocorticoid receptor complex also physically interacts with NF-κB to block its transcriptional activity.\(^{15,16}\) In its inactive state,
NF-κB is sequestered in the cytoplasm by an inhibitory protein named IκB. TNF-α, interleukin-1, microbial pathogens, viral infections, and other inflammatory signals trigger signaling cascades that activate IκB kinases (Fig. 2). Phosphorylation of IκB leads to its ubiquination and degradation by the proteasome, unmasking a nuclear localization signal on NF-κB. In the nucleus, NF-κB binds DNA sequences called NF-κB elements and stimulates the transcription of cytokines, chemokines, cell-adhesion molecules, complement factors, and receptors for these molecules. NF-κB also induces the transcription of cyclooxygenase 2, an enzyme essential for prostaglandin production. Thus, glucocorticoid-induced antagonism of NF-κB and repression of cyclooxygenase 2 is the third mechanism for the inhibition of prostaglandin synthesis after the induction of the antagonists of cPLA₂α, annexin I, and MAPK phosphatase 1 (Fig. 4). Direct interactions between the glucocorticoid receptor and NF-κB probably account for most of the inhibitory effects of glucocorticoids on NF-κB signaling. Despite the analogous nature of glucocorticoid receptor–mediated repression of activator protein 1 and NF-κB, different parts of the surface of the glucocorticoid receptor contact each transcription factor. Glucocorticoids and the glucocorticoid receptor also modulate the activity of other transcription factors.

Recent work suggests that glucocorticoids can have rapid effects on inflammation that are not mediated by changes in gene expression. This best-described nongenomic mechanism involves the activation of endothelial nitric oxide synthetase...
Glucocorticoids stimulate the activity of phosphatidylinositol-3-hydroxykinase (PI3K) in a glucocorticoid receptor–dependent, but transcription-independent, manner in human endothelial cells. Activation of PI3K leads to phosphorylation of Akt. Phosphorylated Akt then phosphorylates and activates eNOS, resulting in the production of nitric oxide. In mice, glucocorticoid-induced acti-
vation of the PI3K–Akt–eNOS pathway protects against ischemia- or reperfusion-induced injury in the heart and the cremaster muscle. This finding is surprising because the production of nitric oxide is generally associated with vasodilation and inflammation. More research is needed to clarify the role of nontranscriptional mechanisms in the inhibition of vasodilation, vascular permeability, and migration of leukocytes across endothelium. Another mechanism of the glucocorticoid-induced inhibition of inflammation involves decreased stability of mRNA for genes for inflammatory proteins such as vascular endothelial growth factor and cyclooxygenase 2. Glucocorticoids clearly act on diverse targets through multiple mechanisms to control inflammation.

**Limitations of Glucocorticoid Therapy**

Although the benefits of glucocorticoid therapy are derived from short-term vascular changes and limited immunosuppression, prolonged or high-dose glucocorticoid therapy has multiple side effects (Table 1). Here, we discuss specific mechanisms involved in a few of these side effects. For instance, extended glucocorticoid treatment can cause hypertension by two distinct mechanisms: one involves renal sodium retention and the ensuing increase in blood volume; a second results from potentiation of vasopressor responses to angiotensin II and catecholamines. Enhanced responses to angiotensin II are due to the induction of angiotensin II receptors by glucocorticoids. Glucocorticoids do not affect the numbers or affinity of α1-adrenergic receptors but, rather, potentiate downstream α1-adrenergic signaling.

Although the systemic vascular resistance induced by glucocorticoids is detrimental, localized changes in vasoreactivity may actually contribute to the beneficial effects of combined treatment with glucocorticoids and β2-agonists in patients with asthma. Specifically, glucocorticoid-enhanced α1-adrenergic signaling (i.e., pulmonary vasoconstriction) could counteract the unfavorable effects of β2-agonists (i.e., pulmonary vasodilation). Patients with asthma have a normal response to β2-agonists in terms of the relaxation of bronchial smooth muscles and increased flow rates, but they have an elevated baseline level of mucosal blood flow that is hypersensitive to vasoconstriction by α1-agonists and insensitive to further vasodilation by β2-agonists. A two-week course of inhaled glucocorticoids decreased baseline perfusion of pulmonary mucosa and restored vascular responsiveness to β2-agonists in patients with asthma. In this study, glucocorticoid modulation of the action of α1-agonists was not determined; thus, it is unclear whether catecholamine signaling was completely restored to normal. This work illustrates one of the potential advantages of inhaled glucocorticoids, which were developed to target lung tissue and thus decrease the adverse effects of systemic delivery. Unfortunately, inhaled glucocorticoids are absorbed by the circulatory system and still cause side effects such as a decreased growth rate in children.

Longitudinal growth in children is a result of the organized proliferation and differentiation of chondrocytes and the subsequent ossification of the extracellular matrix laid down in the growth plates of long bones. Stem cells reside at the epiphyseal end of the growth plate and give rise to proliferating chondrocytes. Moving toward the metaphyseal bone, chondrocytes slow their rate of proliferation, begin to hypertrophy, and produce extracellular matrix proteins and matrix metalloproteinases. As chondrocytes synthesize this scaffolding, they take up calcium and secrete calcium phosphate and hy-

### Table 1. Tissue-Specific Side Effects of High-Dose or Prolonged Glucocorticoid Therapy.

<table>
<thead>
<tr>
<th>Tissue</th>
<th>Side Effects</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adrenal gland</td>
<td>Adrenal atrophy, Cushing’s syndrome</td>
</tr>
<tr>
<td>Cardiovascular system</td>
<td>Dyslipidemia, hypertension, thrombosis, vasculitis</td>
</tr>
<tr>
<td>Central nervous system</td>
<td>Changes in behavior, cognition, memory, and mood (i.e., glucocorticoid-induced psychoses), cerebral atrophy</td>
</tr>
<tr>
<td>Gastrointestinal tract</td>
<td>Gastrointestinal bleeding, pancreatitis, peptic ulcer</td>
</tr>
<tr>
<td>Immune system</td>
<td>Broad immunosuppression, activation of latent viruses</td>
</tr>
<tr>
<td>Integument</td>
<td>Atrophy, delayed wound healing, erythema, hypotrichosis, perioral dermatitis, petechiae, glucocorticoid-induced acne, striae rubrae distensae, telangiectasia</td>
</tr>
<tr>
<td>Musculoskeletal system</td>
<td>Bone necrosis, muscle atrophy, osteoporosis, retardation of longitudinal bone growth</td>
</tr>
<tr>
<td>Eyes</td>
<td>Cataracts, glaucoma</td>
</tr>
<tr>
<td>Kidney</td>
<td>Increased sodium retention and potassium excretion</td>
</tr>
<tr>
<td>Reproductive system</td>
<td>Delayed puberty, fetal growth retardation, hypogonadism</td>
</tr>
</tbody>
</table>

The New England Journal of Medicine
Downloaded from nejm.org at UNIVERSITY OF CHICAGO LIBRARIES on May 8, 2013. For personal use only. No other uses without permission. Copyright © 2005 Massachusetts Medical Society. All rights reserved.
Glucocorticoids slow longitudinal growth by reducing the proliferation of chondrocytes and inducing apoptosis of these cells. Inhibition of insulin-like growth factor I signaling is one mechanism underlying decreased chondrocyte proliferation. In contrast to their effects in the circulatory system, glucocorticoid-induced apoptosis in chondrocytes involves the suppression of signaling through the Akt pathway. Interestingly, insulin-like growth factor I increases the phosphorylation of Akt and acts as a survival factor in glucocorticoid-treated chondrocytes. Although there is generally a period of catch-up growth once glucocorticoid therapy is stopped, sustained treatment with substantive amounts of glucocorticoids during childhood is often associated with decreased adult stature.

Glucocorticoids also have damaging effects on bone in adults. Osteoporosis and an increased risk of fractures are the main side effects of glucocorticoid therapy. Osteoporosis is mediated in part by the binding of glucocorticoid receptors to negative glucocorticoid-responsive elements that inhibit transcription of osteocalcin in osteoblasts; osteocalcin is an important extracellular matrix protein that promotes bone mineralization. Several other side effects of glucocorticoids, including the inhibition of corticotropin-releasing hormone and the expression of pro-opiomelanocortin, are also mediated by negative glucocorticoid-responsive elements (Fig. 4). Glucocorticoids exacerbate osteoporosis by inducing apoptosis in osteoblasts and by increasing the activity of osteoclasts. Some of these effects are directly mediated by glucocorticoid receptors in bone cells, whereas indirect effects are mediated by interactions with other endocrine signals.

The repair of aseptic wounds is also inhibited by glucocorticoids. For example, fractures trigger inflammation and the production of cytokines crucial for the healing and remodeling of bone. In addition to blocking cytokine signaling, glucocorticoids inhibit the synthesis of matrix metalloproteinases and collagen, which are important factors in wound repair. Glucocorticoids also promote gluconeogenesis in the liver, the degradation of proteins to free amino acids in muscle (and muscle atrophy), and lipolysis, ultimately producing hyperglycemia. There are currently no means of ameliorating the side effects of prolonged glucocorticoid therapy that function at the level of the glucocorticoid receptor or the glucocorticoid-responsive elements; rather, treatments such as insulin (or its analogues) for glucocorticoid-induced diabetes, bisphosphonates for osteoporosis, and standard lipid regulators for dyslipidemia are often used. This problem has led to research that has identified potentially selective glucocorticoids.

SELECTIVE GLUCOCORTICOIDs AND FUTURE THERAPY

The pleiotropic effects of glucocorticoids lie between two theoretical extremes. On the one hand, their manifold effects could be inseparable. Alternatively, each effect could be fully dissociated. It has been posited that the antiinflammatory effects of glucocorticoids are primarily mediated by the inhibition of NF-kB and activator protein 1, whereas their side effects result from the activation of transcription. Although this hypothesis is overly simplistic, a recent study described a novel glucocorticoid (ZK216348) with a pattern of repression and activation of transcription that was dramatically different from that of known glucocorticoids. The level of glucocorticoid required to repress interleukin-8 in monocytes was 8 to 12 times as high as that required to induce tyrosine aminotransferase in liver cells. In contrast, the ratio of ZK216348 required to repress interleukin-8 to the level required to activate tyrosine aminotransferase was just 0.4, which was reflected in a better therapeutic index in vivo. Thus, it is possible to develop ligands that inhibit NF-kB–induced expression of inflammatory genes and activate transcription by means of glucocorticoid-responsive elements much more selectively than do currently available glucocorticoids.

Mechanistically, ligands with different structures induce different receptor conformations; for example, the position of helix 12 differs between glucocorticoid receptors bound to agonists and receptors bound to antagonists. Helix 12 closes behind dexamethasone as it sits in the hormone-binding pocket. In this position, helix 12 recruits coactivators required for ligand-dependent transcription. The antagonist mifepristone resides in the same pocket as dexamethasone but causes helix 12 to assume a position that precludes coactivator binding and results in the recruitment of transcriptional corepressors. Further comparison of glucocorticoid analogues reveals that they have divergent transcriptional activities. It is also important that subtle mutations in the GR gene
can be used to differentiate the repression of transcription by activator protein 1 from repression by NF-κB and that different glucocorticoids vary in their capacity to activate genomic and nongenomic mechanisms. These observations highlight the potential for the development of selective glucocorticoids with improved therapeutic profiles.

The rational development of compounds that dissociate the effects of glucocorticoids will require intricate knowledge of the structure of receptors bound to various ligands and an understanding of the way different isoforms of the glucocorticoid receptor activate each signaling pathway. Several commercial entities are actively pursuing these goals. Despite our optimism, it would be naive to suggest that therapeutic effects and side effects are mediated by separate mechanisms or that one could develop ligands that exclusively activate one molecular mechanism. Consequently, it will also be important to optimize the pharmacokinetic and pharmacodynamic properties of new drugs and to develop novel ways to target these drugs to inflamed tissues, as is the case with inhaled glucocorticoids.

**CONCLUSIONS**

The potency of glucocorticoids as inhibitors of diverse inflammatory disorders guarantees their continued use as therapeutic agents. The antiinflammatory and immunosuppressive effects of glucocorticoids rely on several molecular mechanisms, which have been elucidated by basic research. Three main mechanisms include direct effects on gene expression by the binding of glucocorticoid receptors to glucocorticoid-responsive elements (i.e., the induction of annexin I and MAPK phosphatase 1), indirect effects on gene expression through the interactions of glucocorticoid receptors with other transcription factors (i.e., NF-κB and activator protein 1), and glucocorticoid receptor-mediated effects on second-messenger cascades (i.e., the PI3K-Akt–eNOS pathway). Unfortunately, because some of these mechanisms are also involved in physiologic signaling rather than inflammatory signaling, the therapeutic effects of glucocorticoids in inflammation are often accompanied by clinically significant side effects. It is unclear whether isoforms of the glucocorticoid receptor are differentially involved in signaling through each of these mechanisms. Similarly, we do not know whether glucocorticoid-induced activation of certain mechanisms alleviates specific diseases or causes particular side effects. If this sort of signaling specificity exists in vivo, there will be tremendous potential for the development of synthetic ligands that activate antiinflammatory mechanisms but do not affect other pathways. Such drugs would in essence mimic the beneficial effects of natural glucocorticoids without their detrimental side effects.

**REFERENCES**


20. Zhang T, Haws P, Wu Q. Multiple varia-
bles in catons: a mechanism for cell-


23. Webster JC, Oakley RH, Jewell CM, Cidl-
owski JA. Proinflammatory cytokines regu-
late human glucocorticoid receptor gene expression and lead to the accumulation of the dominant negative beta isoform: a mecha-
nism for the generation of glucocorticoid re-


25. Torrego A, Pujol J, Rocas-Jerez J, Mullol J, Xaubet A, Picado C. Glucocorticoid recep-
tor isoforms alpha and beta in in vitro cyto-

26. Yudt MR, Cidlowski JA. Molecular iden-

27. Lu NZ, Cidlowski JA. Translational regu-

28. Ismaili N, Garabedian MJ. Modulation of glucocorticoid receptor function via phos-

29. Wallace AD, Cidlowski JA. Proteasome-
mediated glucocorticoid receptor degrada-

30. Wang X, Pongrac JL, DeFranco DB. Glu-
cocorticoids receptors in hippocampal neu-

31. Le Dearn Y, Mincheneau N, Le Goff P, Michel D. Potentiation of glucocorticoid re-
ceptor transcriptional activity by sumoyla-

32. Holmstrom S, Van Antwerp ME, McGuiz-
Llubi JA. Direct and distinguishable inhibi-

33. Lioniakis MS, Kontoyiannis DP. Gluco-


35. Miller DB, O’Callaghan JP. Neuroendo-
crine aspects of the response to stress. Met-


37. Meduri GU, Yates CR. Systemic inflam-

38. Chilhanza IC. Mechanisms of corticoste-
rone resistance in rheumatoid arthritis: a pu-

39. Tsitoura DC, Rothman PB. Enhance-
ment of MEK/ERK signaling promotes glu-

40. Li LB, Goleva E, Hall CE, Ou LS, Leung DY. Superantigen-induced corticosteroid re-

41. Solito E, de Coupade C, Parente L, Flow-
er RJ, Russo-Marie F. IL-6 stimulates annex-


43. Tanabe T, Toshniwal N. Cyclooxygenase isozymes and their gene structures and ex-
pression. Prostaglandins Other Lipid Medi-

44. Nissen RM, Yamamoto KR. The glucocor-
ticoid receptor inhibitor NIFkappaB by in-
terfering with serine-2 phosphorylation of the RNA polymerase II carboxy-terminal do-

45. Bladh LG, Liden J, Dahlman-Wright K, Reimers M, Nilsson S, Okret S. Identifica-
tion of endogenous glucocorticoid repressed genes differentially regulated by a gluco-
corticoid receptor mutant able to separate hormone and viral factor-kappaB and activa-

46. Barnes PJ, Adcock IM. Transcription fac-
tors and asthma. Eur Respir Rev 1998;12:221-
34.


48. Perretti M, Ahluwalia A. The microcirc-


51. Toh ML, Yang Y, Leech M, Santos L, Mor-
rand EF. Expression of mitogen-activated protein kinase kinases 1, 2, and 3 in rheumatoid arthritis: up-regula-
tion by interleukin-1beta and glucocorti-


